



International Journal of Innovative Pharmaceutical Research

Journal homepage: www.ijipr.com

Method Development and Validation of Sucralfate and Oxetacaine in Bulk Drug and Formulation by Rp-Hplc

Sujitha Parimala S*

Sri Padmavathi School of Pharmacy, Tiruchanoor (Po), Tirupathi, Andhra Pradesh, India-517503.

ABSTRACT

A simple and reliable RP-HPLC chromatographic method was developed for the quantitative determination of Sucralfate and Oxetacaine. Chromatography was carried out by reversed-phase technique on a C₁₈ column with a mobile phase composed of acetonitrile and phosphate buffer (pH 8.9) in the proportion of 30:70 v/v, at a flow rate of 1mL/min with a λ_{\max} of 282 nm. This method was found to be specific and accurate with the mean recovery of 99.60% and 100.32% for Sucralfate and Oxetacaine respectively. The linearity of the proposed method was investigated in the range of 500-1500 $\mu\text{g/mL}$ and 10-30 $\mu\text{g/mL}$ for sucralfate and Oxetacaine respectively with a run time of 20min (Retention time of Sucralfate 3.5min and for Oxetacaine 5.4min). Assay content of Sucralfate and Oxetacaine was determined and the mean assay was found to be 99.93% and 100.2%. The method was also evaluated for robustness, LOD, LOQ and the results obtained were satisfactory. Overall, the proposed method is highly sensitive, precise and accurate and can be used for the reliable quantitation of Sucralfate and Oxetacaine.

Keywords: Sucralfate, Oxetacaine, IR, RP-HPLC, ICH guidelines, Validation.

INTRODUCTION

Sucralfate is a Hexadeca- μ -hydroxytetracosahydroxy [μs -[1, 3, 4, 6-tetra-O-sulfo- β -D-fructofuranosyl- α -D-glucopyranoside tetrakis (hydrogen sulfato) 8-)] hexadecaaluminium. This anti-ulcer drug is a sucrose sulfate-aluminium complex that binds to the mucosa, thus creating a physical barrier that impairs diffusion of hydrochloric acid in the gastrointestinal tract and prevents degradation of mucus by acid (Jensen SL and Funch Jensen P, 1992). Oxetacaine is a 2, 2'-(2-hydroxyethylimino) bis [N-(1, 1-dimethyl-2-phenylethyl)-N-methyl acetamide], which is a local anesthetic drug (Seifter J *et al.*, 1962). It interacts with a receptor situated within the voltage sensitive sodium channel and raises the threshold of channel opening thereby sodium ions permeability fails to increase in response to an impulse or stimulus. This combination is used in the treatment of Peptic ulcers. Sucralfate and Oxetacaine were determined by high performance liquid chromatography with UV detection at 282nm. Linearity for detector response was observed in the concentration range of 500-1500 $\mu\text{g/mL}$ for sucralfate and 10-30 $\mu\text{g/mL}$ for oxetacaine. The aim of this study

was to develop a simple, sensitive precise liquid chromatographic method with PDA detection for the determination of sucralfate and oxetacaine.

MATERIALS AND METHODS

Chemicals and reagents

Dipotassium hydrogen phosphate, Methanol, Acetonitrile were obtained from MERCK Chemicals, Mumbai. Milli Q Water, Distilled Water were procured from In House Production (Lara drugs Private Ltd).

High performance liquid chromatography

Waters HPLC with Empower2 software, 2695 Photodiode Array Detector with data processing capacity was used. A Hypersil BDS column C₁₈, 250 X 4.6mm, 5 μ was used. The p^H measurement was performed by using Labotronics. As a degasser, Eneratek ultrasonicator was used. Typical operating conditions include Flow rate 1mL/min; Injection Volume 10 μL ; Wavelength 282nm; Column Temperature 30^oC; Run time of 20min.

Preparation of solutions

Preparation of solution A

Dissolve 17.418g of Dipotassium hydrogen phosphate in 1000mL of water (pH 8.9). Filter through 0.45 μ or finer porosity membrane filter.

Preparation of solution B

*Corresponding author

Sujitha Parimala S

Email id: sujithanissie7@gmail.com

Take Acetonitrile in 1000mL beaker and degas it to remove air bubbles.

Preparation of mobile phase

Prepare a degassed mixture of solution A & solution B in the ratio of 70:30 v/v.

Preparation of diluent

Prepare a degassed mixture of water & Methanol in the ratio of 70:30 v/v.

Assay Calculation

FORMULA:

$$\frac{AT}{AS} \times \frac{WS}{DS} \times \frac{DT}{WT} \times \frac{Avg.Wt}{LC} \times \text{Purity of STD}$$

$$\% \text{ Assay of Sucralfate} = \frac{382685}{383857} \times \frac{500}{100} \times \frac{5}{25} \times \frac{100}{5000} \times \frac{25}{5} \times \frac{5000}{500} \times 99.2 = 99.93\%$$

$$\% \text{ Assay of Oxetacaine} = \frac{774593}{773580} \times \frac{10}{100} \times \frac{5}{25} \times \frac{100}{5000} \times \frac{25}{5} \times \frac{5000}{500} \times 99.8 = 100.2\%$$

Validation of the developed HPLC method

System suitability

Standard or System Suitability Solution for Sucralfate

Weigh and transfer accurately 500mg of Sucralfate working standard into a 100mL clean, dry volumetric flask, add some amount of diluent and sonicate to dissolve. Make upto the volume with diluent. Dilute 5mL of this solution to 25mL with diluent.

Standard or System Suitability Solution for Oxetacaine

Weigh and transfer accurately 10mg of Oxetacaine working standard into a 100mL clean, dry volumetric flask, add some amount of diluent and sonicate to dissolve. Make upto volume with diluent. Dilute 5mL of this solution to 25mL with the diluent.

Evaluation of system suitability

Inject 10µl of system suitability solution before and after the analysis into the chromatograph and record the chromatograms. The column efficiency as determined from the Sucralfate peak is NLT 2500 USP plate count and USP tailing for the same peak is NMT 2.0. %RSD for peak areas of five injections of standard solution is NMT 2.0.

Precision

The precision of the method was demonstrated through two parameters which are injection reproducibility (system precision) and the method precision.

System precision (Injection reproducibility)

For injection reproducibility, consider five injections of system suitability solution were made and the relative standard deviation for the replicate injections was calculated.

Method Precision

For method precision, 6 individual preparations were made from the same batch and the individual peaks areas were measured. 5g of the suspension is taken in a 100mL volumetric flask. Add some amount of diluent. Sonicate and degas it. Then it is made upto the mark with the diluent. 5mL of the above solution is transferred to a 25mL volumetric flask and made upto the mark with the diluent. Six samples were prepared with a target concentration of about 1000µg/mL from sucralfate and 20µg/mL from oxetacaine.

Accuracy

The accuracy of an analytical method is the closeness of the results obtained by that method to the true value for the sample. It is expressed as recovery (%).The experiment was performed in triplicate for 100%, six times for 50%, and 150%.

6. Limit of Detection

$$\text{LOD of Sucralfate} = 3.3\sigma/S$$

$$= 3.3 \times 25.0066 / 385.4$$

$$= 0.2141 \mu\text{g/mL}$$

$$\text{LOD of Oxetacaine} = 3.3\sigma/S$$

$$= 3.3 \times 538.2 / 7349$$

$$= 0.2416 \mu\text{g/mL}$$

7. Limit of Quantification

$$\text{LOQ of Sucralfate} = 10\sigma/S$$

$$= 0.643 \mu\text{g/mL}$$

$$\text{LOQ of Oxetacaine} = 10\sigma/S$$

$$= 0.7248 \mu\text{g/mL}$$

Figure.1 Standard chromatogram of Sucralfate and Oxetacaine

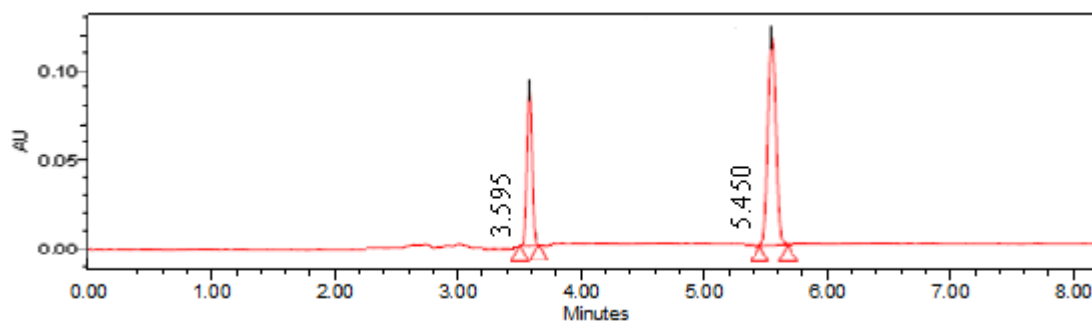


Figure.2 System suitability

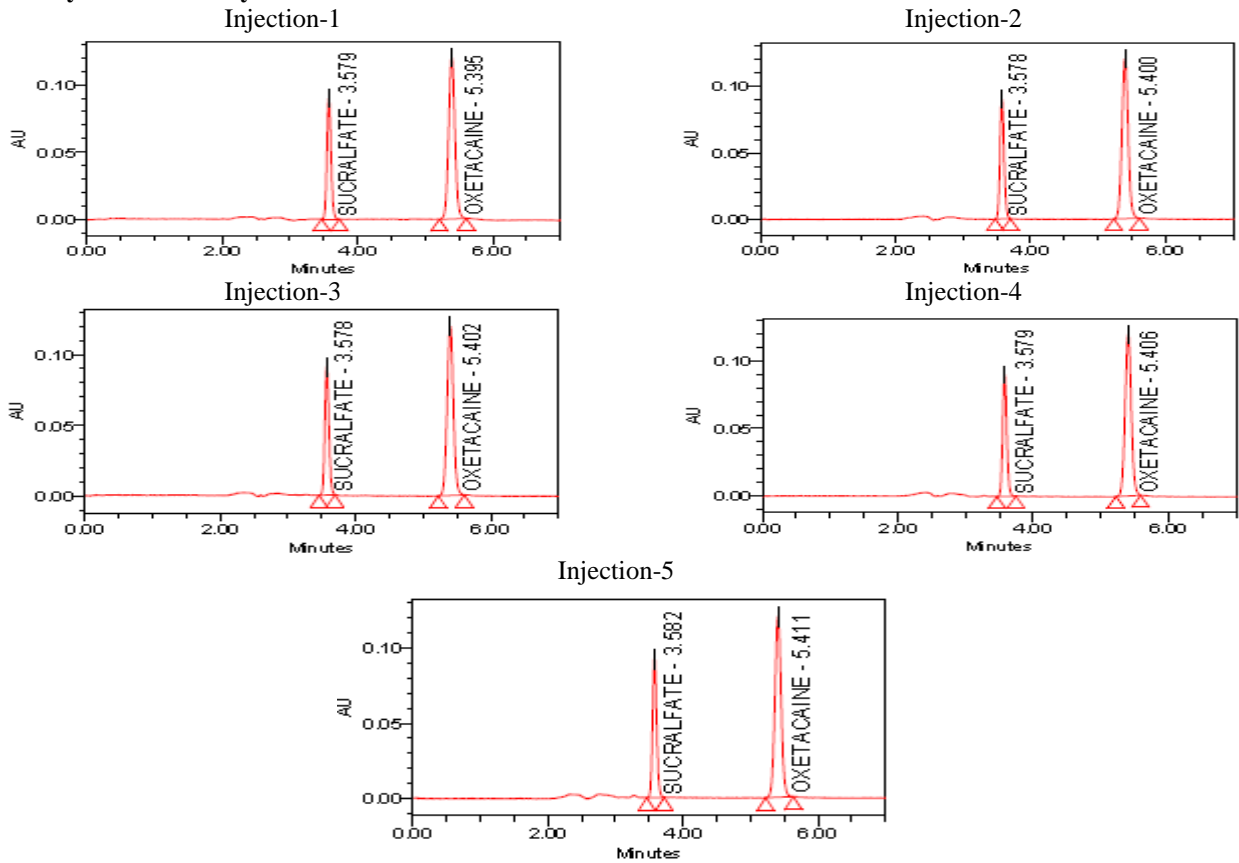


Figure.3 Linearity of sucralfate and oxetacaine

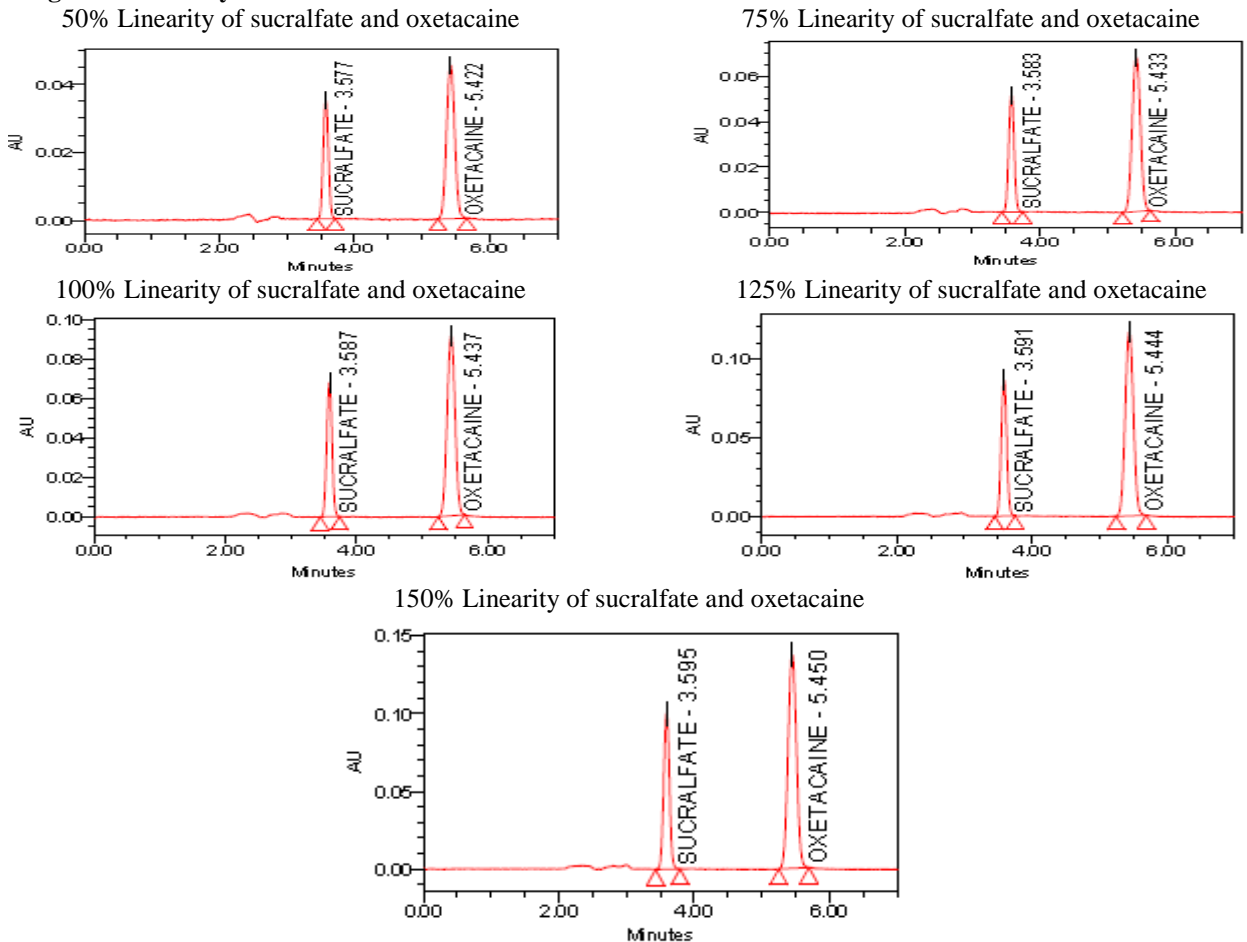


Figure.4 Calibration graph for sucralfate

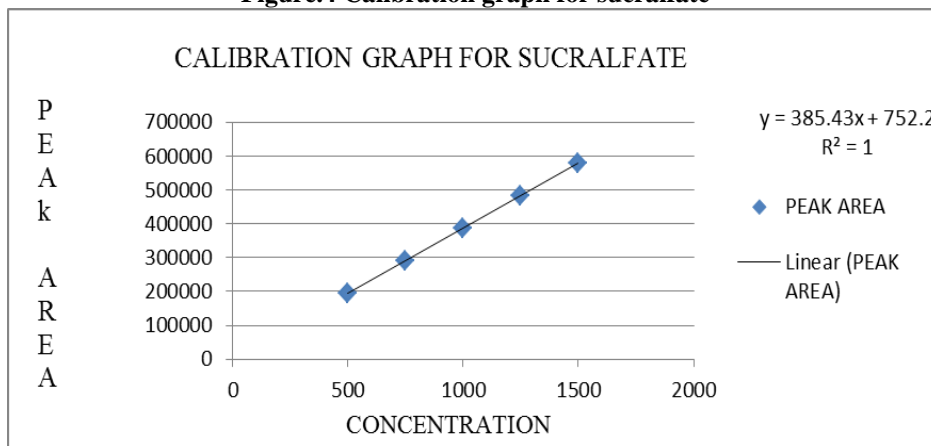


Figure.5 Calibration graph for oxetacaine

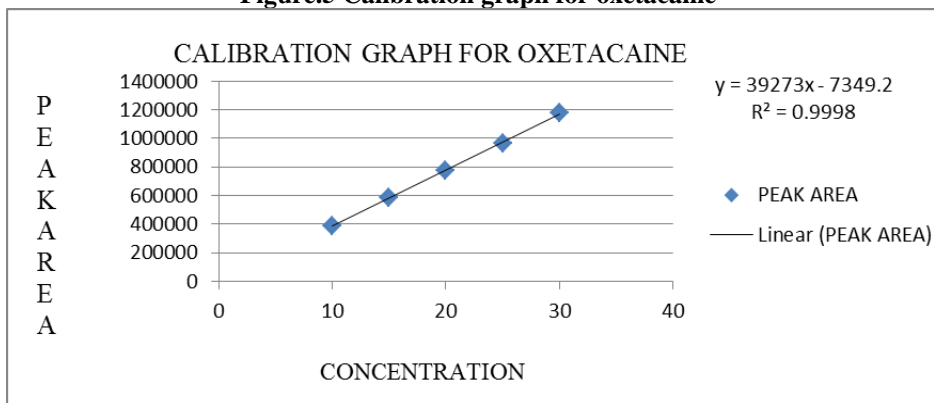


Table.1 System suitability for sucralfate

S. No	Inj	Name	RT	Area	Tailing	Plate count
1	1	SUCRALFATE	3.579	380907	1.128	16691
2	2	SUCRALFATE	3.578	378636	1.122	16965
3	3	SUCRALFATE	3.578	380652	1.118	16518
4	4	SUCRALFATE	3.579	385972	1.126	16437
5	5	SUCRALFATE	3.582	393120	1.129	16587
Mean				383857		
Std. Dev				5842		
%RSD				1.5		

Table.2 System suitability for oxetacaine

S. No	Inj	Name	RT	Area	Tailing	Plate count
1	1	OXETACAINE	5.395	775432	1.050	16288
2	2	OXETACAINE	5.400	771889	1.052	16360
3	3	OXETACAINE	5.402	773217	1.047	16381
4	4	OXETACAINE	5.406	770853	1.033	16380
5	5	OXETACAINE	5.411	776512	1.040	16168
Mean				773580		
Std.Dev				2369		
%RSD				0.3		

Table.3 Linearity of Sucralfate

S. No	Level Solution for Linearity	Concentration of Sucralfate (µg /mL)	Area
1	50%	500	193551
2	75%	750	289706
3	100%	1000	386004
4	125%	1250	482884
5	150%	1500	578744

Table.4 Linearity of Oxetacaine

S. No	Level Solution for Linearity	Concentration of Oxetacaine ($\mu\text{g}/\text{mL}$)	Area
1	50%	10	387960
2	75%	15	582188
3	100%	20	775383
4	125%	25	968339
5	150%	30	1176718

Table.5 Injection reproducibility of sucralfate

S. No	Inj	Name	RT	Area	Tailing	Plate count
1	1	SUCRALFATE	3.579	380907	1.128	16691
2	2	SUCRALFATE	3.578	378636	1.122	16965
3	3	SUCRALFATE	3.578	380652	1.118	16518
4	4	SUCRALFATE	3.579	385972	1.126	16437
5	5	SUCRALFATE	3.582	393120	1.129	16587
Mean				383857		
Std. Dev				5942		
%RSD				1.3		

Table.6 Injection reproducibility of oxetacaine

S. No	Inj	Name	RT	Area	Tailing	Plate count
1	1	OXETACAINE	5.395	775432	1.050	16288
2	2	OXETACAINE	5.400	771889	1.052	16360
3	3	OXETACAINE	5.402	773217	1.047	16381
4	4	OXETACAINE	5.406	770853	1.033	16380
5	5	OXETACAINE	5.411	776512	1.040	16168
Mean				773580		
Std.Dev				2369		
%RSD				0.3		

Table.7 Method precision for sucralfate

S.No	Solution ID	Peak area
1	Preparation-1	383960
2	Preparation-2	379687
3	Preparation-3	383258
4	Preparation-4	381622
5	Preparation-5	383925
6	Preparation-6	383659
SD		1706.353
%RSD		0.445

Table.8 Method precision for oxetacaine

S.No	Solution ID	Peak area
1	Preparation-1	775987
2	Preparation-2	773417
3	Preparation-3	769664
4	Preparation-4	771317
5	Preparation-5	779802
6	Preparation-6	777372
SD		3826.485
%RSD		0.493

Table.9 Method precision and assay of sucralfate

Solution ID	Percentage assay of sucralfate
Preparation-1	99
Preparation-2	98
Preparation-3	99
Preparation-4	99
Preparation-5	99

Preparation-6	99
Mean	99
Standard deviation	0.44
%RSD	0.45

Table.10 Method precision and assay of oxetacaine

Solution ID	Percentage assay of oxetacaine
Preparation-1	100
Preparation-2	100
Preparation-3	99
Preparation-4	100
Preparation-5	101
Preparation-6	100
Mean	100
Standard deviation	0.49

Table.11 Accuracy for sucralfate

S.No	Spiked Level	µg/mL added	µg/mL found	Area	%Recovery	%Mean	%RSD
1.	50%	495.000	486.79	188366	98.01	98	0.4
		495.000	485.08	187702	98		
		495.000	490.73	189888	99		
		495.000	485.29	187785	98.37		
		495.000	484.99	187668	98		
		495.000	486.14	188112	98.05		
2.	100%	990.000	988.95	382678	100	100	0.3
		990.000	983.66	380631	99		
		990.000	988.61	382545	100		
3.	150%	1485.000	1501.60	581047	101	101	0.3
		1485.000	1492.57	577554	101		
		1485.000	1504.47	582157	101		
		1485.000	1499.39	580192	101		
		1485.000	1495.49	578683	101		
		1485.000	1497.81	579580	101		
						Mean %RSD	0.3

Table.12 Accuracy for oxetacaine

S.No	Spiked level	µg/mL added	µg/mL found	Area	%Recovery	%Mean	%RSD
1.	50%	9.900	9.65	374025	97	98	1.3
		9.900	9.62	372888	97		
		9.900	9.87	382657	100		
		9.900	9.89	383242	100		
		9.900	9.58	371296	97		
		9.900	9.74	377401	98		
2.	100%	19.800	20.09	778557	101	101	0.8
		19.800	19.80	767363	100		
		19.800	20.01	775644	101		
3.	150%	29.7	30.38	1177258	102	102	0.1
		29.7	30.37	1177040	102		
		29.7	30.44	1179637	102		
		29.7	30.48	1181389	103		
		29.7	30.41	1178593	102		
		29.7	30.41	1178675	102		
						Mean %RSD	0.73

Robustness**Table.13 Robustness parameters of Sucralfate ($\pm 0.2\text{mL}/\text{min}$)**

S. No	Parameter	Condition	System suitability results for sucralfate			
			RT	Area	USP tailing	Theoretical plates
1	Flow rate by $\pm 0.2\text{mL}/\text{min}$	1.2 mL/min	2.988	382678	1.102	7183
		0.8 mL/min	4.465	380631	1.053	9147

Table.14 Robustness parameters of Sucralfate ($\pm 5^\circ\text{C}$)

S. No	Parameter	Condition	System suitability results for sucralfate			
			RT	Area	USP tailing	Theoretical plates
1	Column Oven temperature by $\pm 5^\circ\text{C}$	35 $^\circ\text{C}$	3.595	389171	1.058	8024
		25 $^\circ\text{C}$	3.581	390722	1.026	8924

Table.15 Robustness parameters of oxetacaine ($\pm 0.2\text{mL}/\text{min}$)

S. No.	Parameter	Condition	System suitability results for Oxetacaine			
			RT	Area	USP tailing	Theoretical plates
1	Flow rate by $\pm 0.2\text{mL}/\text{min}$.	1.2 mL/min	4.536	778557	1.117	7247
		0.8 mL/min	6.788	767363	1.063	9368

Table.16 Robustness parameters of oxetacaine ($\pm 5^\circ\text{C}$)

S. No.	Parameter	Condition	System suitability results for Oxetacaine			
			RT	Area	USP tailing	Theoretical plates
1	Column Oven temperature by $\pm 5^\circ\text{C}$	35 $^\circ\text{C}$	5.450	785501	1.117	7247
		25 $^\circ\text{C}$	5.414	786834	1.063	9368

Summary and conclusion

A simple and reliable RP-HPLC chromatographic method was developed for the quantitative determination of Sucralfate and Oxetacaine. Chromatography was carried out by reverse-phase technique on a C_{18} column with a mobile phase composed of acetonitrile and phosphate buffer (pH 8.9) in the proportion of 30:70 v/v, at a flow rate of 1mL/min with a λ_{max} of 282 nm. This method was found to be specific and accurate with the mean recovery of 99.60%

and 100.32% for Sucralfate and Oxetacaine respectively. The linearity of the proposed method was investigated in the range of 500-1500 $\mu\text{g}/\text{mL}$ and 10-30 $\mu\text{g}/\text{mL}$ for sucralfate and Oxetacaine respectively with a run time of 20min (Retention time of Sucralfate 3.5min and for Oxetacaine 5.4min). Assay content of Sucralfate and Oxetacaine was determined and the mean assay was found to be 99.93% and 100.2%.The method was also evaluated for robustness, LOD, LOQ and the results obtained were satisfactory.

REFERENCES

- Beckett AH, Stenlake JB, Practical pharmaceutical chemistry. 4th Ed, New Delhi, 1997: 275-325.
- Chung Chow Chan, Herman Lam, YC Lee, Xue-Ming Zhang. Analytical Method development and Instrument Performance Verification, John-Wiley & Sons Inc., New York, 2004, 137.
- Donald L Pavia, Gary M Lampman and George S Kriz. Introduction to Spectroscopy, 3rd Ed, Thomson Learning, USA, 2001, 13-101.
- General Chapter 1225, Validation of compendial methods, United States Pharmacopeia 30, National Formulary 25, Rockville, Md., USA, The United States Pharmacopoeial Convention, Inc, 2007.
- Kaur H. Instrumental methods of chemical analysis, High Performance Liquid Chromatography, 2003, 1044-1045.
- Lloyd R Synder, Joseph J Kirkland, Joseph L Glajch. Practical HPLC Method Development, 2nd Ed, Wiley-Interscience, New York, 2007, 1-20.
- Nieman Timothy A, Skoog Douglas A, Holler F James, Pacific Grove CA: Principles of instrumental analysis, 8th Ed, 1998,123.
- Paul C Sadek. The HPLC Solvent Guide, 2nd Ed, Ch.1, Wiley-Interscience, New York, 2002, 1-44.
- Ravi Shankar S. Instrumental methods of chemical analysis, 2001, 18(11).
- Satinder Ahuja and Michael W. Dong, Handbook of Pharmaceutical Analysis by HPLC, 1st Ed., 6, Ch.6, Elsevier Academic Press, UK, 2005, 145-188.
- Sethi PD. HPLC Quantitative Analysis of Pharmaceutical formulations, 1st Ed, CBS Publishers and Distributors, Delhi, 2001, 3-94.
- Sharma BK. Instrumental Methods of Chemical Analysis, 23rd Ed., Goel Publishing House, Meerut, 2004: 68-192.
- Talanta. *History of analytical chemistry*. 1989;36(1-2):1-9.
- Validation of Analytical Procedures: Text and Methodology, Q2 (R1), Step4, ICH Harmonized Tripartite Guidelines 2005.
- Veronika R Meyer. Practical High Performance Liquid Chromatography, 4th Ed., John-Wiley & Sons Inc., New York, 2004, 117.

Willard HH, Merritt LL, Dean JJA, Frank AS. Instrumental methods of analysis, 7th Ed., CBS Publishers and Distributors, New Delhi, 1986,147.
Yuri Kazakevich, Rosario Lubrutto. HPLC for Pharmaceutical Scientists, John-Wiley & Sons, Inc., 2007, 107.